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A Selective 3-Acylation of Tetramic Acids and The First Synthesis of Ravenic Acid

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Abstract: 3-Acyltetramic acids, including delicate 3-oligoenoyl derivatives such as the *Penicillium* metabolite ravenic acid, were prepared in two high-yielding steps. Reaction of tetramic acids with the ylide Ph₃PCCO afforded exclusively the corresponding 3-acylylidene tetramic acids. These were amenable to Wittig olefinations with aliphatic, aromatic, saturated and unsaturated aldehydes after deprotonation with KO^tBu. Due to its simplicity, selectivity and tolerance of pH-sensitive groups this method is superior to the established acylation protocols by Jones and Yoshii. It is also applicable to the synthesis of 3-acyltetronic acids. The new 3-oligoenoyl tetramic acids exhibited structure-dependent antimicrobial and cytotoxic activity.

Introduction

Tetramic acids (i.e., pyrrolidine-2,4-diones) and tetronic acids (i.e., dihydrofuran-2,4-diones) are widespread in nature.^[1-3] They are produced by a variety of marine and terrestrial organisms including bacteria, algae, sponges, fungi and lichens. The 3-acyl substituted derivatives are particularly often associated with biological activity. Various strategies exist for their synthesis. Among these the base-induced Lacey-Dieckmann cyclisation^[4] of *N*-(β-ketoacyl)-α-amino esters is the most widely adopted one,^[5] since directly affording the 3-acyltetramic acids. A potential drawback of this protocol is the frequently observed racemisation at C-5 of the pyrrolidine-2,4-dione core.^[6] An alternative two-step approach first generates the tetramate by condensation of α-amino acids or esters with a dipolar C₂-building block such as Meldrum's acid^[7] or the stable ylide Ph₃PCCO (**1**).^[8] The tetramic acids thus obtained are subsequently acylated at C-3 either with acyl chlorides and BF₃-diethyl etherate acc. to Jones^[9] or with carboxylic acids and DCC/DMAP acc. to Yoshii.^[10] Both acylation methods are tricky. The former is not compatible with acid-sensitive functionalities and skipped carbon-carbon double bonds while the latter tends to fail erratically or to yield 4-*O*-acylated products instead. Herein we report a new selective 3-acylation of tetramic and tetronic acids with ylide **1** and a downstream Wittig alkenation with the so-formed acyl ylides.

Results and Discussion

We had long since observed that tetronic acids, 4-hydroxycoumarins and pyrazol-5-ones reacted with Ph₃PCCO (**1**) under mild, pH-neutral conditions to leave exclusively the corresponding 3-phosphoranylideneacyltetronic acids, 3-phosphoranylideneacyl-4-oxocoumarins, or 4-phosphoranylideneacylpyrazol-5-ones, respectively.^[11] Unfortunately, these products failed to undergo Wittig olefination with aldehydes, a reaction that would provide access for example to the 3-oligoenoyltetronic acid motif occurring in dozens of natural products. X-Ray and NMR studies had revealed π-delocalisation, as well as H-chelate or even phosphonium salt character of the tricarbonyl ylide moiety in these compounds as a reason for their inactivity. Desultory attempts to "switch" them active by removal of the chelated proton with various bases were all unsuccessful. For more systematic studies we now prepared the congenerous 3-acylylidic tetramic acids, e.g. **3**, in virtually quantitative yield by treating the well soluble *N*-Boc-protected tetramic acids **2** with Ph₃PCCO. Next, we identified potassium *tert*-butoxide in THF as a base appropriate for the deprotonation / activation of ylides **3**. The resulting potassium salts were not normally isolated but reacted right away with the respective aldehyde as solutions in THF at reflux. This afforded the corresponding *N*-Boc-3-(α-hydroxydienyl)pyrrolidine-2,4-diones

which were not purified but treated with trifluoroacetic acid (TFA) to liberate the target compounds **4** in yields ranging from 60 to 80%. The same conversion was possible with N-alkyl- and N-H-substituted tetramic acids, e.g. **5**, and with tetronic acids such as **7** (Scheme 1). All steps proceeded with retention of the configuration at C-5 of the starting tetramic or tetronic acids as to HPLC comparison with authentic racemic product samples. It is also worthy of note that the *E*-configured C=C bond introduced in the course of the Wittig alkenation can be removed by catalytic hydrogenation without affecting the formal C=C bond at C-3.

((Scheme 1 here))
((Table 1 here))

The merits of this new acylation protocol are its regioselectivity and mildness of conditions that allow for the presence of acid-sensitive functionalities such as conjugated C=C bonds. We demonstrate this by the first synthesis of ravenic acid **4f** which was originally obtained from a microfungus *Penicillium* sp. (MINAP9902) isolated from the interior of fruiting bodies of the myxomycete *Lycogala epidendrum* collected in south east Queensland. Larger amounts of **4f** were later extracted from culture broths of this fungus. It was found active against methicillin-resistant *Staphylococcus aureus*.^[12] Ylide **3a**, accessible by acylation of N-Boc-pyrrolidine-2,4-dione **2a** (R¹ = H)^[7b] with Ph₃PCCO in 98% yield, was first deprotonated with KO^tBu in THF and then treated with 2-methylocta-2*E*,4*E*,6*E*-trienal **12** to leave ravenic acid **4f** after deprotection with TFA (Scheme 2). HPLC purification eventually afforded an orange crystalline solid in 62% yield with respect to **2a**. Aldehyde **12** was readily prepared. An *E*-selective Horner-Emmons alkenation of sorbinaldehyde with the di-anion of 2-diethoxyphosphorylpropionic acid **10**^[13] gave 2-methylocta-2*E*,4*E*,6*E*-trienoic acid **11** in 72% yield. Acid **11** was treated with SOCl₂ in dichloromethane and the resulting crude acid chloride was immediately reduced with an excess of LiAlH(O^tBu)₃ at -70 °C to afford aldehyde **12** in 50% yield after purification.

((Scheme 2 here))

Table 2 summarizes the results of agar diffusion assays with compounds **4**. In line with the original report^[12] we found antimicrobial activity for ravenic acid **4f** against *Staphylococcus aureus*. It was also active against *Mycobacterium phlei*. Analogue **4a** was equally active against *S. aureus* and even more active than **4f** against *Mycobacterium phlei* and *Micrococcus luteus*. A high degree of unsaturation of the sidechain at C-3 seems not to be a prerequisite for antibiotic activity of 3-acyltetramic acids in these bacteria.

((Table 2 here))

Some derivatives of **4** were also noticeably cytotoxic. For instance, in MTT tests compounds **4a** and **4e** exhibited an IC₅₀(48h) < 15 μM against HL-60 human leukemia cells. With an IC₅₀(48h) value of 22 μM against multidrug-resistant KB-V1 human cervix carcinoma cells compound **4e** even surpassed the efficacy of the clinical anticancer drug doxorubicin. This efficiency is not merely due to the detergent-like nature of compounds such as **4a** and **4e**. For instance, tetramic acid **4a**, while active also against primary human umbilical vein endothelial cells (HUVEC; from Lonza) with an IC₅₀ of 9.3 μM, had a distinct impact on cellular membranes only at much higher concentrations. In hemolysis assays with red blood cells from sheep (from Fiebig Nährstofftechnik, Idstein-Niederau, Germany) we found an ED₅₀ of 250 μM. In contrast, ravenic acid **4f** had little effect both on HUVEC and on erythrocyte membranes

with IC₅₀ and ED₅₀ > 100 μM. A detailed study including more 3-acyltetramic acids as well as tumour cell lines will be disclosed elsewhere.

Conclusion

We have developed a protocol for the synthesis of 3-acyltetramic acids based upon the regioselective C-3-acylation of tetramic acids with the phosphorus ylide Ph₃PCCO. The resulting 3-triphenylphosphoranylideneacyltetramic acids could be deprotonated with potassium *tert*-butoxide affording salts of yet unknown structure which underwent Wittig alkenations with various types of aldehydes. The conditions are mild enough to avoid racemisation of sensitive stereocentres and to allow the introduction of highly unsaturated sidechains at C-3 which are prone to rearrangements and polymerisations under the acidic conditions of the Jones acylation protocol. If undesired, the newly formed C=C bond can be removed selectively by catalytic hydrogenation. The same protocol is applicable to the acylation of tetronic acids. We are currently applying it to the synthesis of more complex natural compounds and we also want to gain structural information on the potassium ylide salt intermediates to better understand the origin of their reactivity.

Experimental Section

General methods: Melting points were recorded on an Electrothermal 9100 apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer Spectrum One FT-IR spectrophotometer equipped with an ATR sampling unit. Nuclear magnetic resonance (NMR) spectra were recorded under conditions as indicated on a Bruker Avance 300 spectrometer. Chemical shifts (δ) are given in parts per million downfield from TMS as an internal standard. Mass spectra were recorded using a Varian MAT 311A (EI). Microanalyses were carried out with a Perkin-Elmer 2400 CHN elemental analyser. For column chromatography Merck silica gel 60 (230-400 mesh) was used. TLC: silica gel 60 F254 (Merck). Optical rotations were recorded at 589 nm with a Perkin-Elmer polarimeter 241. A Knauer system with UV detector K-2000 and pump K-1800 was used for preparative HPLC. Analytical HPLC was performed on a Beckman system with solvent module 126 and a diode array detector 168 equipped with a Nucleodex CD-β-PM column (Macherey-Nagel). Protosil Solvents (HPLC grade) were purchased from Merck. THF was dried over Na/K-alloy and CH₂Cl₂ was dried over P₂O₅. Starting compounds were prepared according to literature procedures or purchased from Fluka, Aldrich or Acros Organic and used without further purification. The weak acid anion exchanger Dowex MPWA was bought from Aldrich.

Synthesis of 3-triphenylphosphoranylideneacyltetramic acids 3 or 6 – general procedure: Under an inert atmosphere a solution of Ph₃PCCO (302 mg, 1.0 mmol) in dry THF (20 mL) was added dropwise over a period of 20 min to a refluxing solution of the respective pyrrolidine-2,4-dione **2** or **5** (1.0 mmol) in dry THF (60 mL). Heating was continued for another 16 h, then half of the solvent was evaporated, pentane was added to the remainder and the product was allowed to precipitate. It was collected by filtration, washed and dried or recrystallised.

1-*tert*-Butoxycarbonyl-3-[(triphenylphosphoranylidene)acetyl]pyrrolidine-2,4-dione (3a): White solid (495 mg, 98%) from 1-*t*-butoxycarbonylpyrrolidine-2,4-dione **2a**^[7b] (199 mg); m.p. 194°C. ¹H NMR (CDCl₃): 2:1 mixture of ylide^a and betaine^b: δ=1.45 (s, 9H; Me₃^b), 1.48 (s, 9H; Me₃^a), 3.77 (s, 2H; 5-H^b), 3.94 (s, 2H; 5-H^a), 5.12 (d, *J*=12.5 Hz, 1H; CH₂P), 5.27 (d, *J*=20.5 Hz, 1H; P=CH), 7.47–7.74 (m, 15H; PPh₃), 12.38 ppm (s, br., 1H; OH); ¹³C NMR (75.5 MHz, CDCl₃): ylide (2 rotamers): δ=27.6 (Me₃), 51.2/54.0 (C-5), 55.7 (d, *J*_{PC}=106.9 Hz; P=CH), 56.3 (d, *J*_{PC}=106.8 Hz; P=CH), 81.3/81.5 (CMe₃), 93.2/95.8 (C-3), 124.1 (d, *J*_{PC}=93.7 Hz; C^{sp2}), 150.7 (CO₂), 168.2 (C-2), 187.6 (C-1'), 191.9 ppm (C-4); betaine: δ=27.5 (Me₃), 35.1 (d; *J*_{PC}=52.8 Hz; P-CH₂), 52.4 (C-5), 81.1 (CMe₃), 103.4 (C-3), 119.8 (d, *J*_{PC}=88.5 Hz; C^{sp2}), 150.3 (CO₂), 172.2 (C-2), 178.7 (C-1'), 189.9 ppm (C-4); further unassignable phenyl signals of both isomers: 128.4, 128.5, 128.6, 129.3, 129.4, 129.7, 129.8, 129.9, 131.8, 131.9, 132.1, 133.0, 133.1, 133.2, 133.3, 133.8, 133.9, 134.4, 134.5 ppm; ³¹P NMR (161.7 MHz, H₃PO₄/ext, CDCl₃): δ=15.6 (ylide), 22.8 ppm (betaine); IR (ATR): ν=1751, 1620, 1557, 1436, 1328, 1153, 1103, 844, 690 cm⁻¹; MS (EI): *m/z* (%): 501 (8) [M⁺], 401 (10), 301 (100), 262 (20) [PPh₃]⁺, 183 (35), 151 (15), 77 (30), 57 (29). Anal. calcd (%) for C₂₉H₂₈NO₃P: C 69.45, H 5.63, N 2.79; found: C 69.51, H 5.66, N 2.83.

(25) $[M]^+$, 241 (10), 149 (10), 126 (42), 44 (100). HRMS (EI): m/z 258.1136 calcd. for $C_{15}H_{18}NO_5^-$, found: 258.1130; 260.1281 calcd. for $C_{15}H_{18}NO_5^+$, found: 260.1287. Anal. calcd (%) for $C_{15}H_{17}NO_3$: C 69.48, H 6.61, N 5.40; found: C 69.50, H 6.66, N 5.48.

(2E,4E,6E)-2-Methylocta-2,4,6-trienoic acid (11) was obtained from 2-diethoxyphosphorylpropionic acid **10** (4.20 g, 20.0 mmol) and sorbinaldehyde (2.10 mL, 20.0 mmol) according to a general literature procedure.^[13b] Recrystallisation from hexane/diethyl ether (2:1) left a white solid of m.p. 125–130°C; yield: 2.17 g (72%). 1H NMR ($CDCl_3$): δ =1.81 (d, J =7.0 Hz, 3H; $CHCH_3$), 1.93 (s, 3H; CH_3), 5.92 (dq, J =15.1, 7.0 Hz, 1H; 7-H), 6.20 (dd, J =15.1, 10.4 Hz, 1H; 6-H), 6.35 (dd, J =14.8, 11.5 Hz, 1H; 4-H), 6.52 (dd, J =14.8, 10.4 Hz, 1H; 5-H), 7.30 ppm (d, J =14.8 Hz, 1H; 3-H); ^{13}C NMR (75.5 MHz, $CDCl_3$): δ =12.8 (Me), 18.7 (Me), 126.7 (C-3), 127.2 (C-2), 133.2 (C-5), 135.1 (C-4), 140.4 (C-6), 141.4 (C-7), 172.1 ppm (C-1); IR (ATR): ν =2999, 2520, 1675, 1600, 1418, 1267, 988, 923, 749 cm^{-1} ; MS (EI): m/z (%) 152 (50) $[M]^+$, 137 (10), 107 (100), 105 (18), 91 (75), 79 (36), 77 (23), 65 (19). HRMS (EI): m/z 152.08373 calcd. for $C_9H_{12}O_2$, found: 152.08370. Anal. calcd (%) for $C_9H_{12}O_2$: C 71.03, H 7.95; found: C 70.88, H 7.89.

(2E,4E,6E)-2-Methylocta-2,4,6-trienal (12): Under an inert atmosphere a mixture of acid **11** (3.04 g, 20.0 mmol), freshly distilled $SOCl_2$ (3.0 mL, 42 mmol), dry CH_2Cl_2 (50 mL) and two drops of dry DMF was stirred at room temperature overnight. All volatiles were removed and the crude acid chloride was re-dissolved in THF (70 mL). The resulting solution was chilled to $-70^\circ C$ and treated dropwise with a 1 M solution of $LiAlH(OtBu)_2$ in THF (1.5 eq, 30 mL, 30.0 mmol). Stirring was continued at this temperature for a further 2 h, then 2 M aq. HCl (25 mL) was added and the mixture was allowed to warm to room temperature. The aqueous layer was extracted with diethyl ether (3 \times 25 mL), the combined organic layers were washed with 2 \times 25 mL each of sat. aq. $NaHCO_3$, sat. aq. NaCl and water. After drying with Na_2SO_4 the solvent was removed and the crude product was purified by column chromatography on silica gel to give aldehyde **12** (1.36 g, 50%) as a colourless oil; R_f =0.44 (hexane/diethyl ether 2:1); 1H NMR ($CDCl_3$): δ =1.81 (d, J =7.0 Hz, 3H; $CHCH_3$), 1.93 (s, 3H; CH_3), 5.94 (dq, J =15.1, 7.0 Hz, 1H; 7-H), 6.21 (dd, J =15.1, 10.4 Hz, 1H; 6-H), 6.53 (dd, J =14.8, 10.4 Hz, 1H; 4-H), 6.81 (dd, J =14.8, 10.4 Hz, 1H; 5-H), 7.31 (d, J =14.8 Hz, 1H; 3-H), 9.41 ppm (s, 1H; CHO); ^{13}C NMR (75.5 MHz, $CDCl_3$): δ =13.8 (Me), 18.6 (Me), 126.8 (C-3), 127.1 (C-2), 131.6 (C-5), 136.0 (C-4), 141.7 (C-6), 148.9 (C-7), 194.5 ppm (C-1); IR (ATR): ν =2927, 2715, 1782, 1675, 1662, 1605, 1240, 1192, 996, 985 cm^{-1} ; MS (EI): m/z (%) 136 (100) $[M]^+$, 121 (40), 107 (36), 93 (62), 91 (78), 77 (58), 65 (26). HRMS (EI): m/z 136.08882 calcd. for $C_9H_{12}O_2$, found: 136.08880. Anal. calcd (%) for $C_9H_{12}O_2$: C 79.37, H 8.88; found: C 79.30, H 8.78.

(S)-3-[(E)-1-Hydroxy-3-(4-methoxyphenyl)allylidene]-5-methylidihydrofuran-2,4-dione (9): Analogously to compounds **4**, lactone **9** was obtained as a yellow solid (180 mg, 66%) from ylide **8** (416 mg, 1.0 mmol) and anisaldehyde (136 μ L); m.p. 97–98°C; $[\alpha]_D^{25}$ -56 (c =0.1, $CHCl_3$). 1H NMR ($CDCl_3$): δ =1.45 (d, J =6.9 Hz, 3H; Me), 3.80 (s, 3H, OMe), 4.57–4.75 (m, 1H; 5-H), 6.86 (d, J =8.9 Hz, 2H; H^a), 7.51 (d, J =15.2 Hz, 1H; 2'-H), 7.58 (d, J =8.9 Hz, 2H; H^b), 7.92 (d, J =15.2 Hz, 1H; 3'-H), 10.45 ppm (s, br., 1H; OH); ^{13}C NMR (75.5 MHz, $CDCl_3$): δ =16.6 (Me), 55.3 (OMe), 81.2 (C-5), 96.3 (C-3), 113.6 (C-2'), 114.5 (C^{exo}), 126.6 (C^{endo}), 131.6 (C^{exo}), 147.7 (C-3'), 163.0 (C^{endo}), 176.6 (C-2), 181.2 (C-1'), 203.9 ppm (C-4); IR (ATR): ν =2933, 1753, 1683, 1624, 1593, 1576, 1512, 1375, 1259, 1171, 1020 cm^{-1} ; MS (EI): m/z (%) 274 (100) $[M]^+$, 245 (12), 201 (50), 174 (73), 131 (40), 77 (30). HRMS: m/z 273.0768 calcd. for $C_{15}H_{18}O_5^-$, found: 273.0763; 275.0914 calcd. for $C_{15}H_{18}O_5^+$, found: 275.0919. Anal. calcd (%) for $C_{15}H_{18}O_5$: C 65.69, H 5.15; found: C 65.73, H 5.15.

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- [1] For reviews on tetramic acids see; a) H.-G. Henning, A. Gelbin, *Adv. Heterocycl. Chem.* **1993**, *57*, 139-185; b) B. J. L. Royles, *Chem. Rev.* **1995**, *95*, 1981-2001; c) E. L. Ghisalberti, in *Studies in Natural Products Chemistry*, Vol. 28/1 (Ed: Atta-ur-Rahman), Elsevier, **2003**, pp. 109-163; d) A. Gossauer, Monopyrrolic natural compounds including tetramic acid derivatives. In *Progress in the Chemistry of Organic Natural Products*, Vol. 86 (Eds: W. Herz, H. Falk, G. W. Kirby), Springer, Wien New York, **2003**, pp. 1-188.
- [2] For reviews on tetronic acids see; a) D. Tejedor, F. Garcia-Tellado, *Org. Prep. Proced. Int.* **2004**, *36*, 35-59; b) A. L. Zografos, D. Georgiadis, *Synthesis* **2006**, 3157-3188.
- [3] For reviews on tetramic and tetronic acids see; a) R. Schobert, *Naturwissenschaften* **2007**, *94*, 1-11; b) R. Schobert, A. Schlenk, *Bioorg. Med. Chem.* **2008**, *16*, 4203-4221.
- [4] R. N. Lacey, *J. Chem. Soc.* **1954**, 850-854.



- [5] a) R. K. Boeckman, C. H. Weidner, R. B. Perni, J. J. Napier, *J. Am. Chem. Soc.* **1989**, *111*, 8036; b) L. A. Paquette, D. MacDonald, L. G. Anderson, J. Wright, *J. Am. Chem. Soc.* **1989**, *111*, 8037; c) S. V. Ley, S. C. Smith, P. R. Woodward, *Tetrahedron* **1992**, *48*, 1145-1174; d) Y. Iwata, N. Maekawara, K. Tanino, M. Miyashita, *Angew. Chem.* **2005**, *117*, 1556-1560; *Angew. Chem. Int. Ed.* **2005**, *44*, 1532-1536; e) L. Burke, D. Dixon, S. V. Ley, F. Rodríguez, *Org. Lett.* **2000**, *2*, 3611-3613; f) A. C. Hart, A. J. Phillips, *J. Am. Chem. Soc.* **2006**, *128*, 1094-1095; g) R. Böhme, G. Jung, E. Breitmaier, *Helv. Chim. Acta* **2005**, *88*, 2837-2841.
- [6] J. Poncet, P. Jouin, B. Castro, L. Nicolas, M. Boutar, A. Gaudemer, *J. Chem. Soc. Perkin Trans. 1* **1990**, 611.
- [7] a) P. Jouin, B. Castro, D. Nisato, *J. Chem. Soc. Perkin Trans. 1* **1987**, 1177-1182; b) S. Hamilakis, D. Kontonassios, C. Sandris, *J. Heterocyclic Chem.* **1996**, *33*, 825-829; c) B. Q. Li, R. W. Franck, *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2629-2634; d) Z. Liu, X. Ruan, X. Huang, *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2505-2507.
- [8] a) R. Schobert, C. Jagusch, C. Melanophy, G. Mullen, *Org. Biomol. Chem.* **2004**, *2*, 3524-3529; b) R. Schobert, C. Jagusch, *Tetrahedron* **2005**, *61*, 2301-2307; c) R. Schobert, M. Dietrich, G. Mullen, J.-M. Urbina-Gonzalez, *Synthesis* **2006**, 3902-3914; d) B. Biersack, R. Diestel, C. Jagusch, G. Rapp, F. Sasse, R. Schobert, *Chem. Biodiv.* **2008**, *5*, 2423-2430.
- [9] a) H. Kohl, S. V. Bhat, J. R. Patell, N. M. Ghandi, J. Nazareth, P. V. Divekar, N. J. de Souza, H. G. Berscheid, H.-W. Fehlhaber, *Tetrahedron Lett.* **1974**, *15*, 983-986; b) R. C. F. Jones, M. J. Begley, G. E. Peterson, S. Sumaria, *J. Chem. Soc. Perkin Trans. 1* **1990**, 1959-1968.
- [10] K. Hori, M. Arai, K. Nomura, E. Yoshii, *Chem. Pharm. Bull.* **1987**, *35*, 4368-4371.
- [11] R. Schobert, S. Siegfried, M. Nieuwenhuyzen, W. Milius, F. Hampel, *J. Chem. Soc. Perkin Trans. 1* **2000**, 1723-1730.
- [12] A. P. Michael, E. J. Grace, M. Kotiw, R. A. Barrow, *J. Nat. Prod.* **2002**, *65*, 1360-1362.
- [13] a) P. Coutrot, A. Ghribi, *Synthesis* **1986**, 661; b) P. Coutrot, A. Ghribi, *Synthesis* **1986**, 790-792.
- [14] M. Storgaard, F. Zaragoza Doerwald, B. Peschke, D. Tanner, *J. Org. Chem.* **2009**, *74*, 5032-5040.
- [15] M. Hosseini, H. Kringelum, A. Murray, J. E. Tonder, *Org. Lett.* **2006**, *8*, 2103-2106.
- [16] R. Schobert, R. Stehle, H. Walter, *Tetrahedron* **2008**, *64*, 9401-9407.

((Scheme and figure legends)):

Scheme 1. Selective 3-acylation of tetronic and tetramic acids.

Scheme 2. Synthesis of ravenic acid **4f**.

Table 1. 3-Alkenoyltetramic acids **4** by acylation with Ph₃PCCO (**1**) / aldehydes

compounds 4	R ¹	R ²	Yield [%]	made via ylide
4a	H	(CH ₂) ₈ Me	76	3a
4b	H	<i>p</i> (C ₆ H ₄)OMe	84	3a
4c	H		78	3a
4d ^[a]	(5 <i>S</i>)- <i>p</i> (OH)Bn		66	3b
4e	(5 <i>S</i>)-CH ₂ Ph	<i>p</i> (C ₆ H ₄)OMe	62	6

[a] precursors **2b** and **3b**: R¹ = (5*S*)-CH₂-*p*(C₆H₄)OtBu.

Table 2. Antibiotic activity^[a] of tetramic acids **4** against selected Gram-positive bacteria^[b]

	4a	4c	4e	4f
<i>Staphylococcus aureus</i>	11	0	0	11
<i>Micrococcus luteus</i>	8	0	0	0
<i>Mycobacterium phlei</i>	11	8	9	9

[a] Agar plates inoculated with the respective microorganisms were incubated with 6 mm cellulose discs containing 20 μL of a methanolic solution (1 mg mL⁻¹) of the compounds tested. The diameters (in mm) of the resulting growth-inhibition zones were determined after 24 h of incubation at 30°C and are cited here. [b] None of the compounds inhibited the growth of the Gram-negative bacteria *E. coli* tolC and *Klebsiella pneumoniae*. Compounds **4b**, **4d** and **9** were inactive in all tested bacteria.

((Text for the Table of Contents)):

Two double bonds from one ylide: Tetramic and tetronic acids react with Ph_3PCCO to give exclusively and quantitatively the corresponding 3-acyl ylides. Once deprotonated with KO^tBu they undergo *E*-selective Wittig reaction with aliphatic and aromatic aldehydes. Even delicate 3-oligoenoyl tetramic acids such as the natural antibiotic ravenic acid are accessible in good yields. New biological properties are reported.

Keywords: tetramic acids • ravenic acid • acylation • antibiotics • phosphorus ylides