

Genome Sequence of Strain MOLA814, a Proteorhodopsin-Containing Representative of the *Betaproteobacteria* Common in the Ocean

Alicia Courties,^{a,b} Thomas Riedel,^{a,b} Michael Jarek,^c Laurent Intertaglia,^{a,b} Philippe Lebaron,^{a,b} Marcelino T. Suzuki^{a,b}

UPMC Université Paris 6, UMR 7621, Observatoire Océanologique, Banyuls-sur-Mer, France^a; CNRS, UMR 7621, LOMIC, Observatoire Océanologique, Banyuls-sur-Mer, France^b; Helmholtz-Centre for Infection Research, Research Group Genomic Analytics, Braunschweig, Germany^c

Strain MOLA814 is a marine betaproteobacterium that was isolated from seawater in the Beaufort Sea. Here, we present its genome sequence and annotation. Genome analysis revealed the presence of a proteorhodopsin-encoding sequence together with its retinal-producing pathway, indicating that this strain might generate energy by using light.

Received 8 November 2013 Accepted 15 November 2013 Published 19 December 2013

Citation Courties A, Riedel T, Jarek M, Intertaglia L, Lebaron P, Suzuki MT. 2013. Genome sequence of strain MOLA814, a proteorhodopsin-containing representative of the *Betaproteobacteria* common in the ocean. *Genome Announc.* 1(6):e01062-13. doi:10.1128/genomeA.01062-13.

Copyright © 2013 Courties et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Marcelino T. Suzuki, suzuki@obs-banyuls.fr.

Marine strain MOLA814 was isolated from a depth of 3 m in the Canadian Beaufort Sea (71°40.294'N, 130°43.674'W). This strain belongs to the *Betaproteobacteria*, and its 16S rRNA sequence is 98% identical to that of strain OTU126, which is described as the 40th most abundant operational taxonomic unit (OTU) among >45,000 sequences from surface ocean planktonic prokaryotes (1).

The genomic DNA of strain MOLA814 was extracted using the cetyltrimethylammonium bromide (CTAB) protocol (2). Library preparation for whole-genome sequencing was performed using the TruSeq DNA PCR-free sample preparation kit (Illumina, San Diego, CA) with 550-bp insert sizes, according to the manufacturer's protocol. Genomic DNA was sheared using a Covaris S2 system (Covaris, Woburn, MA) and subjected to end repair, purification, and ligation of the fragments with multiple indexed adapters for library preparation. Quality control of the prepared library was validated using quantitative PCR (qPCR) (Kapa library quantification kit; Kapa Biosystems, Woburn, MA) and an Agilent Bioanalyzer high-sensitivity (HS) chip (Agilent Technologies, Santa Clara, CA) according to the manufacturers' instructions. Genome sequencing was performed to 250 cycles in both directions in a MiSeq system (Illumina), which generated 2,446,022 total reads (611.5 Mbp). DNA-Seq reads were converted to Fastq format and *de novo* assembled with Velvet 1.2.07 (3). The sequencing data were controlled for general quality features using the FastqMcf tool of ea-utils (<http://code.google.com/p/ea-utils>). The resulting 3 scaffolds with 87× average coverage of the genome were annotated using Prokka 1.7 (4).

The draft genome sequence of strain MOLA814 is 2,859,706 bp in size, contains 2,683 coding sequences, 3 rRNAs, and 39 tRNAs, and has a G+C content of 53.6%.

Interestingly, the genome analysis of strain MOLA814 revealed the presence of a proteorhodopsin-encoding gene sequence (PR) and a putative retinal-producing biosynthetic pathway (5–7). The PR-encoding sequence codes for a green light-absorbing PR-opsin (8, 9) of 263 amino acid residues with the typical features necessary for proton pump activity, like Asp97 and Glu108 resi-

dues (eBAC31A08 numbering). These act as proton acceptor and donor in the retinylidene Schiff base transfer during the PR photocycle. BLAST analysis (10) revealed high PR protein sequence identities to the PR sequences of representatives belonging to the alphaproteobacterial SAR116 clade, like “*Candidatus* Puniceispirillum marinum” IMCC1322 (11).

The presence of a PR-encoding sequence together with its retinal-producing pathway in the genome sequence indicates a putative photoheterotrophic lifestyle. In addition, the genome sequence of strain MOLA814 provides a good opportunity for studying the physiological and ecological functions of a commonly occurring marine betaproteobacterium living in ocean waters.

Nucleotide sequence accession numbers. The whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. [AYMW000000000](https://www.ncbi.nlm.nih.gov/nuccore/AYMW000000000). The version described in this paper is version [AYMW010000000](https://www.ncbi.nlm.nih.gov/nuccore/AYMW010000000).

ACKNOWLEDGMENTS

This work was supported by the French ANR (project RHOME0). Growth and DNA extraction were performed using the Marine Biodiversity and Biotechnology (Bio2Mar) platform of the OOB.

We sincerely thank the Genome Analytics staff (Helmholtz-Centre for Infection Research) for rapidly and efficiently sequencing the genome.

REFERENCES

1. Yooshep S, Neelson KH, Rusch DB, McCrow JP, Dupont CL, Kim M, Johnson J, Montgomery R, Ferriera S, Beeson K, Williamson SJ, Tovchigrechko A, Allen AE, Zeigler LA, Sutton G, Eisenstadt E, Rogers YH, Friedman R, Frazier M, Venter JC. 2010. Genomic and functional adaptation in surface ocean planktonic prokaryotes. *Nature* 468:60–66.
2. Ausubel FM, Brent R, Kingston RE, Moore DD, Smith JA, Seidman JG, Struhl K. 1987. *Current protocols in molecular biology*. John Wiley and Sons, New York, NY.
3. Zerbino DR, Birney E. 2008. Velvet: algorithms for the *de novo* short read assembly using the Bruijn graphs. *Genome Res.* 18:821–829.
4. Victorian Bioinformatics Consortium. 2013. Prokka 1.7. Victorian Bioinformatics Consortium, Clayton, Victoria, Australia. <http://www.vicbioinformatics.com/software/prokka.shtml>.
5. Bèjà O, Aravind L, Koonin EV, Suzuki MT, Hadd A, Nguyen LP,

- Jovanovich SB, Gates CM, Feldman RA, Spudich JL, Spudich EN, DeLong EF. 2000. Bacterial rhodopsin: evidence for a new type of phototrophy in the sea. *Science* 289:1902–1906.
6. Gómez-Consarnau L, González JM, Coll-Lladó M, Gourdon P, Pascher T, Neutze R, Pedrós-Alió C, Pinhassi J. 2007. Light stimulates growth of proteorhodopsin-containing marine *Flavobacteria*. *Nature* 445:210–213.
 7. Kimura H, Young CR, Martinez A, DeLong EF. 2011. Light-induced transcriptional responses associated with proteorhodopsin-enhanced growth in a marine *Flavobacterium*. *ISME J.* 5:1641–1651.
 8. Man D, Wang W, Sabehi G, Aravind L, Post AF, Massana R, Spudich EN, Spudich JL, Bèjà O. 2003. Diversification and spectral tuning in marine proteorhodopsins. *EMBO J.* 22:1725–1731.
 9. Fuhrman JA, Schwalbach MS, Stingl U. 2008. Proteorhodopsins: an array of physiological roles? *Nat. Rev. Microbiol.* 6:488–494.
 10. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J. Mol. Biol.* 215:403–410.
 11. Oh HM, Kwon KK, Kang I, Kang SG, Lee JH, Kim SJ, Cho HC. 2010. Complete genome sequence of “*Candidatus Puniceispirillum marinum*” IMCC 1322, a representative of the SAR116 clade in the *Alphaproteobacteria*. *J. Bacteriol.* 192:3240–3241.