COMMENTARY

Commentary on "Antimicrobial Capacity of Plant Polyphenols against Gram-positive Bacteria: A Comprehensive Review" authored by Enrique Barrajón-Catalán, Institute of Molecular and Cell Biology (IBMC), Miguel Hernandez University (UMH), Avda. Universidad s/n, Elche 03202. Spain

Wolf-Rainer Abraham¹

¹Helmholtz Center for Infection Research, Chemical Microbiology, Inhoffenstrasse 7, 38124 Braunschweig, Germany

We are facing the emergence and extensive spread of antimicrobial resistance of many pathogens against commercial antibiotics [1]. In addition to this, most microorganisms protect themselves by forming biofilms where cells are glued together by a highly structured, self-produced matrix consisting of a complex mixture of polysaccharides, proteins, lipids and DNA [2]. In biofilms cells are not only well protected but they also produce a number of pathogenicity factors and become much more resistant against antibiotics [3] or the immune system of the host. Hence, pathogens organized in biofilms are very difficult to eradicate [4, 5].

Because i) the resistances against many antibiotics are increasing, ii) less and less new antibiotics are brought into the market, and iii) antibiotics are not effective against biofilm forming pathogens alternatives are urgently needed to control bacterial infections [6, 7]. Especially Gram-positive bacteria with their multiresistances are a challenge in the clinics. One strategy of searching for novel drugs is to look for solutions developed by other organisms. As all multicellular organisms are challenged by pathogenic microorganisms forming biofilms on their body they developed a multitude of compounds for their control [8]. For example, several medicinal plants from Cameroon displayed broad bacteriostatic activities [9], a sponge colonizing Penicillium sp. forms the chlorinated polyketide, Hypoxylon fragiformis produces sclerin acting against Staphylococcus aureus biofilm formation [10] or carolactone from the myxobacterium Sorangium cellulosum damages Streptococcus mutans biofilms [11], only to mention a few compounds.

That plants are a source for antimicrobials is known for centuries [12], therefore, Álvarez-Martínez and coworkers reviewed the activity of polyphenols from plants against Gram-positive bacteria [13]. This is a really challenging task because a huge chemical diversity of polyphenolic natural products from plants is known and many of these compounds have multiple targets in the cells acting as antioxidants, anticancer drugs, antivirals, etc. besides being antimicrobials. On top of this several antimicrobial natural products have been reported to have several targets in the cells reducing the risk that pathogens develop resistance [14]. The authors were aware of these problems and tackled them by a consequent organization of their review. They limited their review to Gram-positive bacteria but included both planktonic and sessile cells. With this, their scope was much broader than a similar but shorter review only on the antibiofilm activity of polyphenols [15]. The authors nicely compiled various reports on the antibacterial activities of a number of polyphenols. Unfortunately, the current research focus seems to be on Staphylococcus aureus and its various multiresistant strains, e. g. MRCA. This is also mirrored in the review but the authors tried to broaden the spectrum of antimicrobial activities against Gram-positive bacteria wherever it was possible. They also dealt very well with the "grey zone" of reports were ill characterized mixtures of polyphenols have been tested against Gram-positive bacteria. As from the broad diversity of chemical structures can be expected polyphenols from plants have several modes of action. Most screenings for antibiofilm compounds focus on the communication of the bacterial cells to form a biofilm, the quorum sensing

cascade. This, however, misses all the other targets which control biofilm formation and dispersion [16]. The authors were aware of this bias and, where available, they compiled the different mechanisms offering insights into the many ways for the control of Grampositive bacteria. Such an overview over the mechanisms is very valuable, on the one hand for the set-up of sophisticated screening assays but also on the other hand to broaden the antimicrobial spectrum of bioactive natural compounds.

In my opinion the complex biofilm communities of the oral cavity is a hot topic but not much has been said on dental biofilms where Gram-positive bacteria have a leading role [17]. Not all members of these communities, however, are pathogenic. Some of them may even have a protective role for the teeth. The aim is therefore to control these biofilms and to suppress its pathogenic members. For the control a number of polyphenols have been applied either as pure compounds or as plant extracts [18]. The field of dental biofilms is a nice example where functional food [19], traditional medicine [20, 21] and modern health care [22] meet.

I would like to emphasize another aspect in the search for the optimal antimicrobial polyphenol which was also not much discussed due to space limitation. This is the analyses of the antimicrobial activities of a series of compounds and their connection with the substitution pattern and physical characteristics [23, 24]. These structure activity relationships can lead to improved drugs for the control of pathogens or biofilm infections [25]. Here natural products chemistry meets combinatorial chemistry for the development of natural products for clinical applications [26, 27]. Finally, one should be aware that most of these polyphenols from plants possess only weak to moderate antimicrobial activities. Combining them with commercial antibiotics may boost their activity and may even be effective in biofilm infections [28]. Using combination of different polyphenols, however, requires tight control of their ratios for optimal biological activities as has been mirrored by the variability of results from studies using crude extracts [29]. This faces the problem of standardization of plant extracts both in storage and processing [30, 31].

REFERENCES

Tacconelli, E.; Carrara, E.; Savoldi, A.; Harbarth, S.; Mendelson, M.; Monnet, D.L.; Pulcini, C.; Kahlmeter, G.; Kluytmans, J.; Carmeli, Y.; Ouellette, M.; Outterson, K.; Patel, J.; Cavaleri, M.; Cox, E.M.; Houchens, C.R.; Grayson, M.L.; Hansen, P.; Singh, N.; Theuretzbacher, U.; Magrini, N. WHO pathogens priority list working group.

- Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. Lancet Infect. Dis., 2018, 18(3), 318-327. http://dx.doi.org/10.1016/S1473-3099(17)30753-3 PMID: 29276051
- Costerton, J.W.; Stewart, P.S.; Greenberg, E.P. Bacterial [2] biofilms: a common cause of persistent infections. Science, **1999**, 284(5418), 1318-1322. http://dx.doi.org/10.1126/science.284.5418.1318 PMID: 10334980
- Townsley, L.; Shank, E.A. Natural-product antibiotics: cues [3] for modulating bacterial biofilm formation. Trends Microbiol., 2017, 25(12), 1016-1026. http://dx.doi.org/10.1016/j.tim.2017.06.003 PMID: 28688575
- Malone, M.; Bjarnsholt, T.; McBain, A.J.; James, G.A.; Stoodley, P.; Leaper, D.; Tachi, M.; Schultz, G.; Swanson, T.; Wolcott, R.D. The prevalence of biofilms in chronic wounds: a systematic review and meta-analysis of published data. J. Wound Care, 2017, 26(1), 20-25. http://dx.doi.org/10.12968/jowc.2017.26.1.20 PMID: 28103163
- [5] Arciola, C.R.; Campoccia, D.; Montanaro, L. Implant infections: adhesion, biofilm formation and immune evasion. Nat. Rev. Microbiol., 2018, 16(7), 397-409. http://dx.doi.org/10.1038/s41579-018-0019-y PMID: 29720707
- Martens, E.; Demain, A.L. The antibiotic resistance crisis, [6] with a focus on the United States. J. Antibiot. (Tokyo), 2017, 70(5), 520-526. http://dx.doi.org/10.1038/ja.2017.30 PMID: 28246379
- Abraham, W-R. Controlling biofilms of gram-positive [7] pathogenic bacteria. Curr. Med. Chem., 2006, 13(13), 1509-1524.
 - http://dx.doi.org/10.2174/092986706777442039 PMID: 16787201
- [8] Wright, G.D. Opportunities for natural products in 21st century antibiotic discovery. Nat. Prod. Rep., 2017, 34(7), 694http://dx.doi.org/10.1039/C7NP00019G PMID: 28569300
- [9] Voukeng, I.K.; Beng, V.P.; Kuete, V. Antibacterial activity of six medicinal Cameroonian plants against Gram-positive and Gram-negative multidrug resistant phenotypes. BMC Complement. Altern. Med., 2016, 16(1), 388. http://dx.doi.org/10.1186/s12906-016-1371-y PMID: 27724917
- Yuyama, K.T.; Chepkirui, C.; Wendt, L.; Fortkamp, D.; Stadler, M.; Abraham, W-R. Bioactive compounds produced by Hypoxylon fragiforme against Staphylococcus aureus biofilm. Microorganisms, 2017, 5(4), 80. http://dx.doi.org/10.3390/microorganisms5040080 PMID: 29231891
- Jansen, R.; Irschik, H.; Huch, V.; Schummer, D.; Steinmetz, H.; Bock, M.; Schmidt, T.; Kirschning, A.; Müller, R. Carolacton-A macrolide ketocarbonic acid that reduces biofilm formation by the caries-and endocarditis-associated bacterium Streptococcus mutans. Eur. J. Org. Chem., 2010, 2010, 1284-1289. http://dx.doi.org/10.1002/ejoc.200901126
- Chandra, H.; Bishnoi, P.; Yadav, A.; Patni, B.; Mishra, A.P.; Nautiyal, A.R. Antimicrobial resistance and the alternative resources with special emphasis on plant-based antimicrobials – a review. *Plants (Basel)*, **2017**, *6*(2), 16. http://dx.doi.org/10.3390/plants6020016 PMID: 28394295

- [13] Álvarez-Martínez, F.J.; Barrajón-Catalán, E.; Encinar, J.A.; Rodríguez-Díaz, J.C.; Micol, V. Antimicrobial capacity of plant polyphenols against Gram-positive bacteria: a comprehensive review. *Curr. Med. Chem.*, 2018. http://dx.doi.org/10.2174/0929867325666181008115650 PMID: 30295182
- [14] Brown, E.D.; Wright, G.D. Antibacterial drug discovery in the resistance era. *Nature*, 2016, 529(7586), 336-343. http://dx.doi.org/10.1038/nature17042 PMID: 26791724
- [15] Slobodníková, L.; Fialová, S.; Rendeková, K.; Kováč, J.; Mučaji, P. Antibiofilm activity of plant polyphenols. *Molecules*, 2016, 21(12), 1717. http://dx.doi.org/10.3390/molecules21121717 PMID: 27983597
- [16] Abraham, W-R. Going beyond the control of quorumsensing to combat biofilm infections. *Antibiotics (Basel)*, 2016, 5(1), 3. http://dx.doi.org/10.3390/antibiotics5010003 PMID: 27025518
- [17] Larsen, T.; Fiehn, N.E. Dental biofilm infections an update. APMIS, 2017, 125(4), 376-384. http://dx.doi.org/10.1111/apm.12688 PMID: 28407420
- [18] Ferrazzano, G.F.; Amato, I.; Ingenito, A.; Zarrelli, A.; Pinto, G.; Pollio, A. Plant polyphenols and their anti-cariogenic properties: a review. *Molecules*, 2011, 16(2), 1486-1507. http://dx.doi.org/10.3390/molecules16021486 PMID: 21317840
- [19] Esteban-Fernández, A.; Zorraquín-Peña, I.; de Llano, D.G.; Bartolomé, B.; Moreno-Arribas, M.V. The role of wine and food polyphenols in oral health. *Trends Food Sci. Technol.*, 2017, 69A, 118-130. http://dx.doi.org/10.1016/j.tifs.2017.09.008
- [20] Kawarai, T.; Narisawa, N.; Yoneda, S.; Tsutsumi, Y.; Ishi-kawa, J.; Hoshino, Y.; Senpuku, H. Inhibition of Strepto-coccus mutans biofilm formation using extracts from Assam tea compared to green tea. *Arch. Oral Biol.*, 2016, 68, 73-82. http://dx.doi.org/10.1016/j.archoralbio.2016.04.002 PMID: 27107380
- [21] Karygianni, L.; Al-Ahmad, A.; Argyropoulou, A.; Hellwig, E.; Anderson, A.C.; Skaltsounis, A.L. Natural antimicrobials and oral microorganisms: a systematic review on herbal interventions for the eradication of multispecies oral biofilms. *Front. Microbiol.*, 2016, 6, 1529. http://dx.doi.org/10.3389/fmicb.2015.01529 PMID: 26834707
- [22] Van der Weijden, F.A.; Van der Sluijs, E.; Ciancio, S.G.; Slot, D.E. Can chemical mouthwash agents achieve plaque/gingivitis control? *Dent. Clin. North Am.*, 2015, 59(4), 799-829. http://dx.doi.org/10.1016/j.cden.2015.06.002 PMID: 26427569
- [23] Andrade-Ochoa, S.; Nevárez-Moorillón, G.V.; Sánchez-Torres, L.E.; Villanueva-García, M.; Sánchez-Ramírez, B.E.; Rodríguez-Valdez, L.M.; Rivera-Chavira, B.E. Quantitative structure-activity relationship of molecules constituent of different essential oils with antimycobacterial activity

- against Mycobacterium tuberculosis and Mycobacterium bovis. *BMC Complement. Altern. Med.*, **2015**, *15*(1), 332. http://dx.doi.org/10.1186/s12906-015-0858-2 PMID: 26400221
- [24] Xie, Y.; Yang, W.; Tang, F.; Chen, X.; Ren, L. Antibacterial activities of flavonoids: structure-activity relationship and mechanism. *Curr. Med. Chem.*, 2015, 22(1), 132-149. http://dx.doi.org/10.2174/0929867321666140916113443 PMID: 25245513
- [25] Manner, S.; Skogman, M.; Goeres, D.; Vuorela, P.; Fallarero, A. Systematic exploration of natural and synthetic flavonoids for the inhibition of Staphylococcus aureus biofilms. *Int. J. Mol. Sci.*, 2013, 14(10), 19434-19451. http://dx.doi.org/10.3390/ijms141019434 PMID: 24071942
- [26] Harvey, A.L.; Edrada-Ebel, R.; Quinn, R.J. The reemergence of natural products for drug discovery in the genomics era. *Nat. Rev. Drug Discov.*, 2015, 14(2), 111-129. http://dx.doi.org/10.1038/nrd4510 PMID: 25614221
- [27] Rodrigues, T.; Reker, D.; Schneider, P.; Schneider, G. Counting on natural products for drug design. *Nat. Chem.*, 2016, 8(6), 531-541. http://dx.doi.org/10.1038/nchem.2479 PMID: 27219696
- [28] Estrela, A.B.; Abraham, W.R. Combining biofilmcontrolling compounds and antibiotics as a promising new way to control biofilm infections. *Pharmaceuticals (Basel)*, 2010, 3(5), 1374-1393. http://dx.doi.org/10.3390/ph3051374 PMID: 27713308
- [29] Baptista, A.; Gonçalves, R.V.; Bressan, J.; Pelúzio, M.D.C.G. Antioxidant and antimicrobial activities of crude extracts and fractions of cashew (Anacardium occidentale L.), Cajui (Anacardium microcarpum), and pequi (Caryocar brasiliense C.): a systematic review *Oxidat Med Cell Longevity*, 2018. http://dx.doi.org/10.1155/2018/3753562 PMID: 29849888
- [30] Ghasemzadeh, A.; Jaafar, H.Z.; Rahmat, A. Variation of the phytochemical constituents and antioxidant activities of Zingiber officinale var. rubrum Theilade associated with different drying methods and polyphenol oxidase activity. *Molecules*, 2016, 21(6), 780. http://dx.doi.org/10.3390/molecules21060780 PMID: 27322227
- [31] Kunle, O.F.; Egharevba, H.O.; Ahmadu, P.O. Standardization of herbal medicines A review. *Int. J. Biodivers. Conserv.*, 2012, 4, 101-112. http://dx.doi.org/10.5897/IJBC11.163

Wolf-Rainer Abraham

Helmholtz Center for Infection Research, Chemical Microbiology, Inhoffenstrasse 7, 38124 Braunschweig, Germany;

E-mail: wolf-rainer.abraham@helmholtz-hzi.de

DISCLAIMER: The above article has been published in Epub (ahead of print) on the basis of the materials provided by the author. The Editorial Department reserves the right to make minor modifications for further improvement of the manuscript.